# No membrane? No problem: Engineering the ASGPR receptor for soluble assay development

Bradley Peter<sup>1</sup>, Bo Peng<sup>1</sup>, Alice Ghidini<sup>2</sup>, Anders Gunnarsson<sup>2</sup> and Jianming Liu<sup>1</sup>

- <sup>1</sup>Discovery Biology, Discovery Sciences, R&D, AstraZeneca, Gothenburg, Sweden
- <sup>2</sup>Mechanistic and Structural Biology, Discovery Sciences, R&D, AstraZeneca, Gothenburg, Sweden

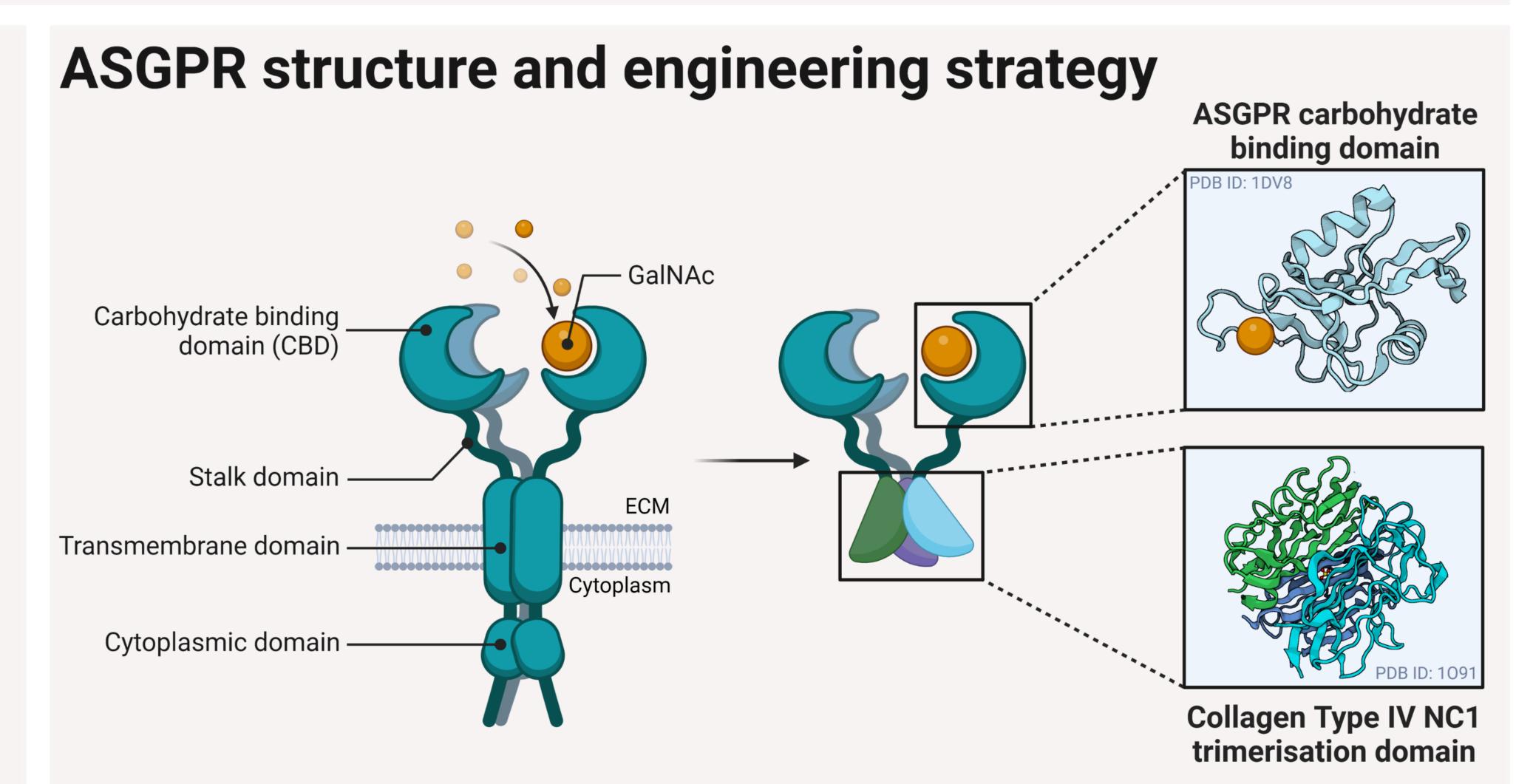


#### Introduction

Asialoglycoprotein receptor (ASGPR) is a trimeric transmembrane receptor that can be exploited for the targeted delivery of therapeutic oligonucleotides to the liver via its natural ligand N-acetyl-D-galactosamine (GalNAc). Developing novel ligands for liver-targeted delivery forms part of the ongoing AstraZeneca Oligonucleotide Platform Build.

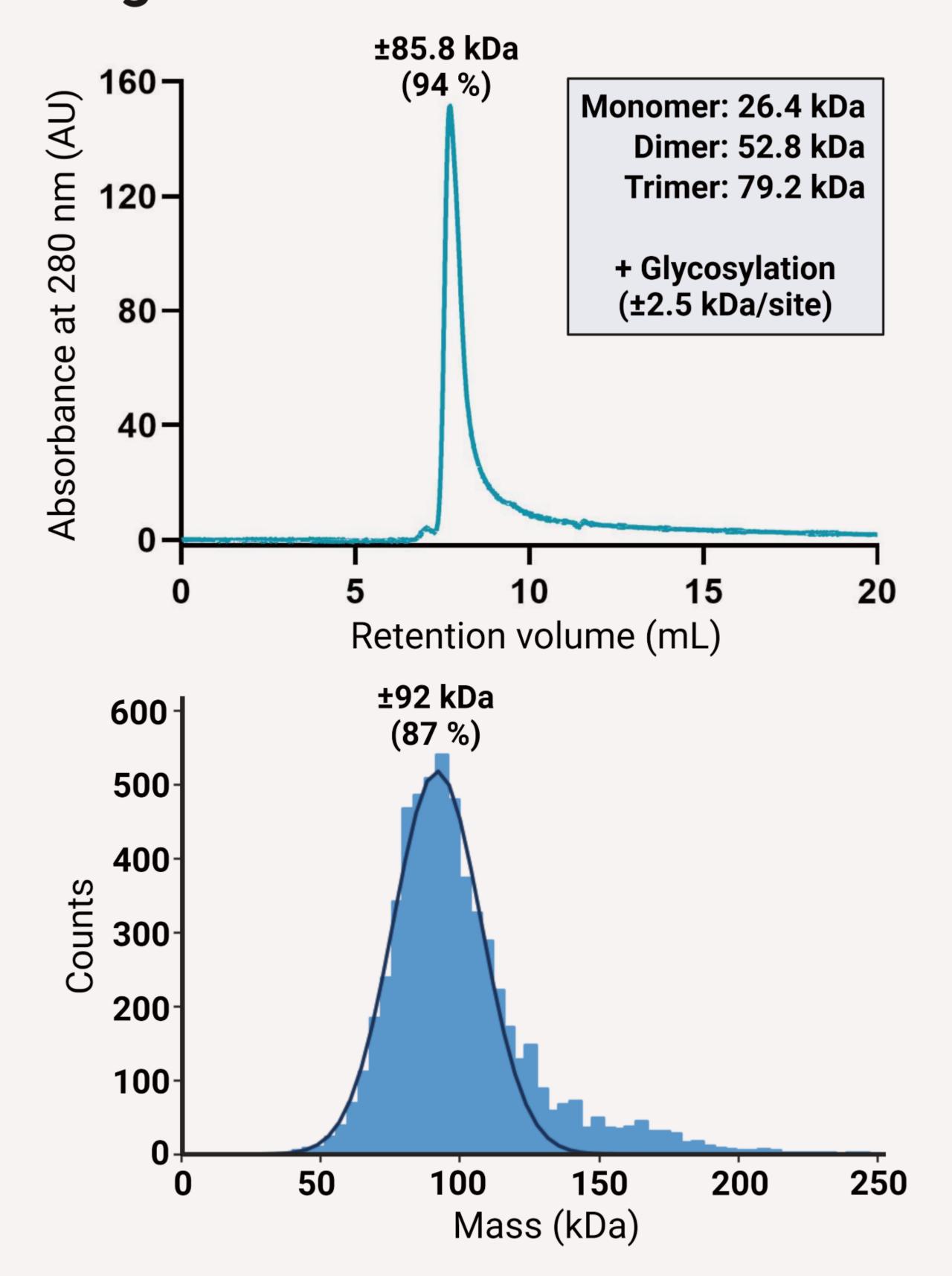
In vitro studies of ASGPR have been limited to the monomeric carbohydrate binding domain (CBD). The absence of a membrane precludes analysis of the trimeric form of the receptor, which is expected to have a higher affinity for trivalent ligands targeting the CBD.

Here, we designed an artificial trimeric construct in the absence of a membrane and comprising the soluble CBD of ASGPR fused to the NC1 trimerisation domain of collagen.



**FIGURE 1**: ASGPR is a heterotrimeric receptor which is internalised following GalNAc binding. Here, we isolated the CBD and engineered a trimerisation domain from collagen onto its N-terminus to promote oligomerisation in the absence of a membrane

### **Engineered ASGPR forms trimers**



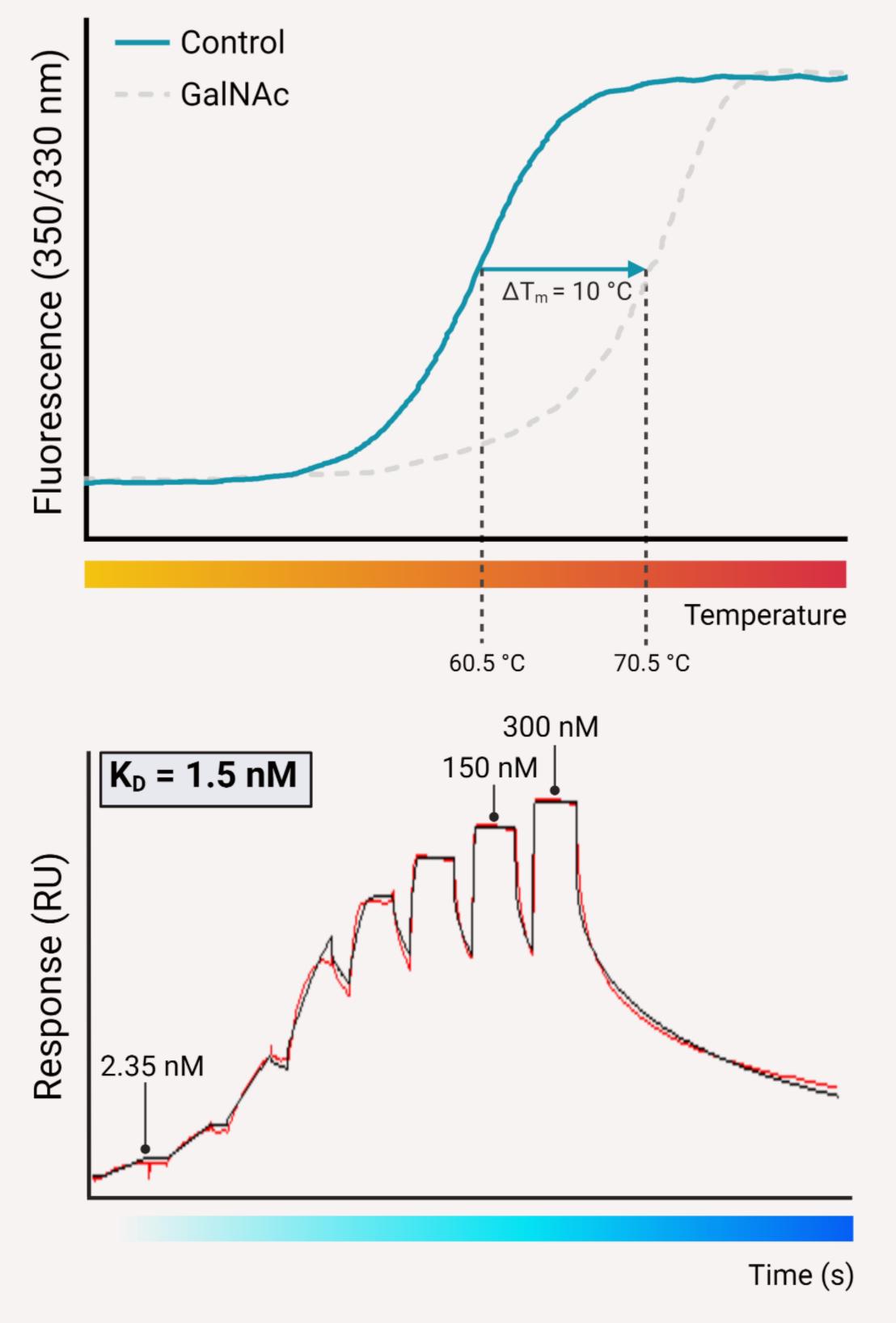
**FIGURE 2**: Engineered ASGPR forms stable trimers when analysed by SEC-MALS (A) and mass photometry (B). The apparent higher  $M_{\rm w}$  compared to the theoretical trimer is due to the presence of three glycosylation sites (+ 7.5 kDa)

#### Conclusions

This approach represents a different way of tackling membrane receptor drug discovery, with the benefits of a native-like soluble oligomer gained whilst avoiding the potential challenges associated with membrane protein purification.

Our next step is to use the Tri-ASGPR construct to generate a high-throughput assay for compound screening.

## Tri-ASGPR is binding-competent



**FIGURE 3**: Engineered ASGPR is capable of binding the native GalNAc substrate. A large thermal shift was observed upon GalNAc binding ( $\Delta T_m = 10$  °C) using nanoDSF (A). Binding was also confirmed using SPR (B), measuring an apparent K<sub>D</sub> of 1.5 nM

## Acknowledgements

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Figures generated using BioRender.

Contact: bradley.peter@astrazeneca.com